

## Myricananone and Myricananadiol: Two New Cyclic ‘Diarylheptanoids’ from the Roots of *Myrica nana*

by Ya-Fang Yu<sup>a)</sup>), Qing Lu<sup>a)</sup>), Li Guo<sup>b)</sup>), Ren-Qiang Mei<sup>a)</sup>), Heng-Xing Liang<sup>a)</sup>), Du-Qiang Luo<sup>a)</sup>), and Yong-Xian Cheng<sup>\*a)</sup>

<sup>a)</sup> State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming, 650204, P. R. China

(phone: +86-871-5223048; fax: +86-871-5223048; e-mail: yxcheng@mail.kib.ac.cn)

<sup>b)</sup> College of Pharmacy, Chengdu University of Traditional Chinese Medicine, Chengdu 610075, P. R. China

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Two new cyclic ‘diarylheptanoids’, myricananone (**1**) and myricananadiol (**2**), were isolated from the roots of *Myrica nana*, together with the known compounds myricanol (**3**), myricanone (**4**), and porson (**5**). Their structures were determined by spectroscopic methods, including 1D- and 2D-NMR as well as HR-ESI-MS analyses.

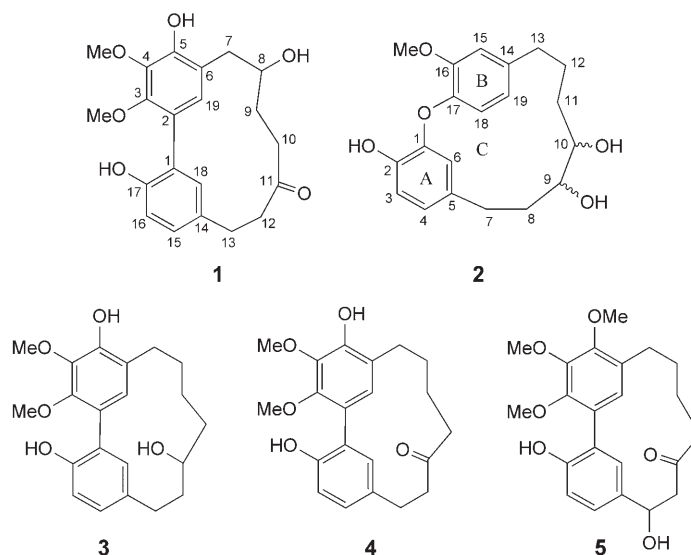
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**Introduction.** – *Myrica nana* CHEVAL. (Myricaceae) is an evergreen shrub native to Yunnan and Guizhou Provinces of mainland China [1]. Its fruits are edible and beneficial for dyspepsia. Its roots are used as a Chinese folk medicine for the treatment of bleeding, diarrhea, stomach pain, burns, and skin diseases [2]. Several triterpenoids, flavonoids, tannins and ‘diarylheptanoids’ have been isolated from other Myricaceae plants [3–11], and some phenolic compounds were previously reported from the fresh leaves of *M. nana* [12]. However, the information on the chemical constituents of the roots of this plant is still scarce.

Herein, we report on the constituents of the 95% EtOH extract of the roots of *M. nana*, from which the cyclic compounds **1**–**5** were isolated. Compounds **1** and **2**, named myricananone and myricananadiol, respectively, are new natural products, and the known compounds **3**–**5** were isolated for the first time from this plant. In this paper, we describe the isolation and structural elucidation of these isolates.

**Results and Discussion.** – Compound **1** was obtained as a colorless, optically inactive powder. Its HR-ESI mass spectrum showed the quasi-molecular  $[M + Na]^+$  ion peak at  $m/z$  395.1467 (calc. 395.1471), corresponding to the molecular formula  $C_{21}H_{24}O_6$ , with ten degrees of unsaturation. Analysis of the <sup>1</sup>H- and <sup>13</sup>C-NMR data of **1** (Table) indicated a cyclic ‘diarylheptanoid’ closely related to the known compounds **3**–**5**. The high similarity of the <sup>13</sup>C-NMR signals of **1** and **3** in the aromatic region indicated that they shared the same diphenyl substitution pattern.

The positions of the OH and C=O groups were determined by <sup>1</sup>H,<sup>1</sup>H-COSY and HMBC experiments (Fig. 1). Two spin systems were readily identified as H–C(7)/H–C(8)/H–C(9)/H–C(10) and as H–C(12)/H–C(13) by the <sup>1</sup>H,<sup>1</sup>H-COSY inter-



actions<sup>1)</sup>. The connection of C(10) and C(12) *via* a C=O group in 11-position was established by the HMBC correlations of CH<sub>2</sub>(9), CH<sub>2</sub>(10), CH<sub>2</sub>(12), and CH<sub>2</sub>(13) with C(11). The linkage of the aliphatic chain with the diphenyl moiety was assembled with the aid of HMBC correlations between CH<sub>2</sub>(7) and C(5), C(6), and C(19); between CH<sub>2</sub>(12) and C(14); between CH<sub>2</sub>(13) and both C(14) and C(15), and between C(13) and both H–C(15) and H–C(18).

In the NOESY spectrum of **1** (Fig. 1), key NOEs were observed between H–C(19) and H–C(7), H–C(8), H<sub>a</sub>–C(9), and H–C(18); between H–C(8) and H<sub>a</sub>–C(10); between H<sub>b</sub>–C(9) and H<sub>b</sub>–C(10); between H–C(18) and H<sub>a</sub>–C(9), H–C(12), and

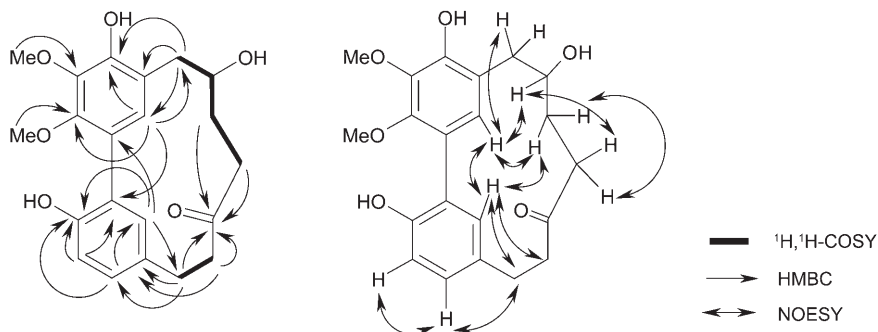


Fig. 1. Key <sup>1</sup>H,<sup>1</sup>H-COSY, HMBC, and NOESY interactions of **1**

<sup>1)</sup> Arbitrary atom numbering.

Table.  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR Data of **1** and **2**. At 400 and 100 MHz, resp.,  $\delta$  in ppm,  $J$  in Hz.

Position <sup>1)</sup>	<b>1</b> <sup>a)</sup>		Position <sup>1)</sup>	<b>2</b> <sup>b)</sup>	
	$\delta(\text{H})$	$\delta(\text{C})$		$\delta(\text{H})$	$\delta(\text{C})$
1		126.8	1		144.8
2		124.1	2		149.3
3		149.0	3	6.73 ( <i>d</i> , $J = 8.0$ )	116.5
4		141.1	4	6.57 ( <i>dd</i> , $J = 8.0, 1.9$ )	122.8
5		150.2	5		133.8
6		123.4	6	5.72 ( <i>d</i> , $J = 1.9$ )	114.5
7a	2.90–3.04 ( <i>m</i> )	36.9	7	2.48–2.62 ( <i>m</i> )	29.4
7b	2.64–2.75 ( <i>m</i> )				
8	4.24 ( <i>m</i> )	71.0	8	1.50–1.54 ( <i>m</i> )	35.2
9a	2.14–2.18 ( <i>m</i> )	32.3	9	2.87–2.92 ( <i>m</i> )	74.9
9b	1.61–1.65 ( <i>m</i> )				
10a	2.64–2.75 ( <i>m</i> )	42.2	10	2.48–2.62 ( <i>m</i> )	76.6
10b	2.90–3.04 ( <i>m</i> )				
11		216.0	11a	1.22–1.24 ( <i>m</i> )	32.9
			11b	1.38–1.47 ( <i>m</i> )	
12a	2.90–3.04 ( <i>m</i> )	43.4	12a	1.63–1.73 ( <i>m</i> )	27.5
12b	2.75–2.81 ( <i>m</i> )		12b	1.90–1.95 ( <i>m</i> )	
13	2.90–3.04 ( <i>m</i> )	29.0	13a	2.48–2.62 ( <i>m</i> )	36.0
			13b	2.87–2.92 ( <i>m</i> )	
14		133.0	14		141.5
15	6.98 (br. <i>d</i> , $J = 8.0$ )	129.6	15	6.98 ( <i>d</i> , $J = 1.7$ )	117.7
16	6.74 ( <i>d</i> , $J = 8.0$ )	117.3	16		152.7
17		152.9	17		144.6
18	6.56 (br. <i>s</i> )	134.2	18	7.08 ( <i>d</i> , $J = 7.8$ )	125.2
19	6.40 ( <i>s</i> )	130.8	19	6.94 ( <i>dd</i> , $J = 7.8, 1.7$ )	122.2
OMe	3.78 ( <i>s</i> )	61.6	OMe	3.64 ( <i>s</i> )	56.4
OMe	3.88 ( <i>s</i> )	61.5	OMe	7.67 ( <i>s</i> )	
			9-OH	3.24 (br. <i>s</i> )	
			10-OH	3.18 (br. <i>s</i> )	

<sup>a)</sup> In  $\text{CD}_3\text{OD}$ . <sup>b)</sup> In  $(\text{D}_6)\text{Acetone}$ .

H–C(13); and between H–C(13) and H–C(15). Since the compound showed no optical activity, it was most likely present as a racemic mixture.

From the above data, the structure of the new compound **1** was, thus, deduced as 3,12,15-trihydroxy-16,17-dimethoxytricyclo[12.3.1.1<sup>2,6</sup>]nonadeca-1(18),2(19),3,5,14,16-hexaen-9-one, and given the trivial name *myricananone*.

Compound **2** was obtained as a colorless, optically active powder ( $[\alpha]_{\text{D}}^{23.6} = -71.4$ ). The molecular formula  $\text{C}_{20}\text{H}_{24}\text{O}_5$  was derived by HR-ESI-MS ( $m/z$  367.1517 ( $[M + \text{Na}]^+$ ; calc. 367.1521), requiring nine degrees of unsaturation. The  $^{13}\text{C}$ -NMR spectrum of **2** (Table) exhibited 20 signals, which were ascribable to two benzene rings, five  $\text{CH}_2$  and two oxygenated CH groups, as well as one MeO function.

The  $^1\text{H}$ -NMR spectrum of **2** showed two *ABX*-type spin systems in the aromatic region, resonating at  $\delta(\text{H})$  7.08 (*d*,  $J = 7.8$  Hz, 1 H), 7.04 (br. *d*,  $J = 7.8$  Hz, 1 H), and 7.06 (br. *s*, 1 H); and at 6.73 (*d*,  $J = 8.0$  Hz, 1 H), 6.57 (*dd*,  $J = 8.0, 1.9$  Hz, 1 H), and

5.72 ( $d$ ,  $J = 1.9$  Hz, 1 H), respectively. The unusual upfield-shifted aromatic signal at  $\delta(\text{H})$  5.71 was assumed to be caused by the anisotropy of the neighboring benzene  $B$ -ring; similar phenomena were observed before in the related structures of galeon and acerogenin B [13][14].

The above data suggested that **2** was a diphenyl-ether analogue of a ‘diaryl-heptanoid’. The substitution patterns of the two benzene rings were resolved on the basis of HMBC correlations (Fig. 2) between H–C(3) and both C(4) and C(5); between H–C(6) and C(1), C(2), and C(4); between 2-OH and both C(2) and C(3); between H–C(15) and C(16); between H–C(18) and C(16), C(17), and C(19); between H–C(19) and C(15); and between 16-MeO and C(16). The positions of the two OH groups in the aliphatic chain were determined as C(9) and C(10), respectively, as derived from the  $^1\text{H}, ^1\text{H}$ -COSY interactions for CH<sub>2</sub>(7)/CH<sub>2</sub>(8)/H–C(9)/H–C(10)/CH<sub>2</sub>(11)/CH<sub>2</sub>(12)/CH<sub>2</sub>(13), for HO–C(9)/H–C(9), and for HO–C(10)/H–C(10), as well as based on the HMBC correlations between H–C(4) and C(7); between H–C(6) and C(7); between H–C(7) and C(4), C(5), and C(6); between H–C(8) and C(5); between H–C(12) and C(14); and between H–C(13) and C(14), C(15), and C(19).

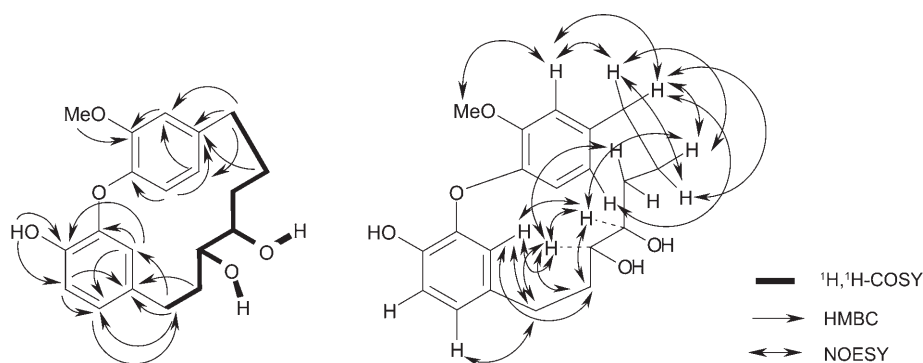


Fig. 2. Key  $^1\text{H}, ^1\text{H}$ -COSY, HMBC, and NOESY interactions of **2**

The unusual upfield shifts of H–C(9) ( $\delta(\text{H})$  2.87–2.92) and H–C(10) ( $\delta(\text{H})$  2.48–2.62) indicated that these two H-atoms were somewhat shielded by the aromatic  $B$ -ring, which can happen when rings  $A$  and  $B$  are almost perpendicular to one another. In addition, key NOEs were observed in the ROESY spectrum of **2** (Fig. 2) between CH<sub>2</sub>(7) and H–C(4), H–C(6), and H–C(9); between CH<sub>2</sub>(8) and H–C(6), H–C(9), and H–C(10); between H–C(6) and both H–C(9) and H–C(10); between H<sub>a</sub>–C(11) and H–C(9); between H<sub>a</sub>–C(12) and H–C(10); and between H–C(19) and H<sub>b</sub>–C(13). This indicated that the flexible ring  $C$  adopts a preferential conformation in solution, in which free rotation about the single bonds is restricted.

Thus, from the above data, the structure of the new compound **2** was deduced as 17-methoxy-2-oxatricyclo[13.2.2.1<sup>3,7</sup>]icosa-1(17),3(20),4,6,15,18-hexaene-4,10,11-triol, and given the trivial name *myricananadiol*. The relative and absolute configurations at the two stereogenic centers, C(9) and C(10), remain to be determined.

The three known cyclic ‘diarylheptanoids’ were identified as myricanol (**3**) [9], myricanone (**4**) [9], and porson (**5**) [15], respectively, by comparison of their spectroscopic data with literature values. These compounds were all isolated for the first time from *M. nana*.

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### Experimental Part

**General.** Column chromatography (CC): silica gel (200–300 mesh, 10–40  $\mu\text{m}$ ; *Qingdao Marine Chemical Factory*, China),  $C_{18}$  reverse-phase (RP) silica gel (40–63  $\mu\text{m}$ ; *Daiso Co.*, Japan), and *Sephadex LH-20* (*Amersham Pharmacia Biotech*, Sweden). Thin-layer chromatography (TLC): silica gel *GF<sub>254</sub>* (10–40  $\mu\text{m}$ ; *Qingdao*). All solvents were distilled before use. UV/VIS Spectra: *Shimadzu UV-2401PC* spectrophotometer;  $\lambda_{\text{max}}$  in nm. Optical rotations: *JASCO-20C* digital polarimeter.  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR Spectra: *Bruker AM-400* spectrometer, at 400 and 100 MHz, resp; chemical shifts  $\delta$  in ppm rel. to  $\text{Me}_4\text{Si}$ ,  $J$  in Hz.  $^1\text{H}$ ,  $^1\text{H}$ -COSY, HMQC, HMBC, and NOESY Spectra: *Bruker DRX-500* spectrometer. MS: *VG AutoSpec-3000* mass spectrometer; in  $m/z$ . HR-ESI-MS: *API QSTAR Pulsar-1* mass spectrometer.

**Plant Material.** The roots of *Myrica nana* CHEVAL. were collected from Songhua dam, a Kunming suburb in Yunnan Province, China, in July 2005, and were identified by Dr. Y. M. Shui, Kunming Institute of Botany. A voucher specimen (CHYX0391) was deposited at the State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming, P. R. China.

**Extraction and Isolation.** The dried and powdered roots of *M. nana* (7.0 kg) were extracted with 95% EtOH at reflux ( $3 \times$ ). The combined extracts were concentrated *in vacuo*. The residue was suspended in  $\text{H}_2\text{O}$ , and then extracted with AcOEt. The resulting AcOEt-soluble extract (276 g) was subjected to CC ( $\text{SiO}_2$ ;  $\text{CHCl}_3/\text{MeOH}$  5:1) to afford four fractions (*Fr. 1* – *Fr. 4*). *Fr. 1* (67 g) was purified by CC ( $\text{SiO}_2$ ;  $\text{CHCl}_3/\text{AcOEt}$  4:1) to yield *Fr. 1.1* – *Fr. 1.3*. *Fr. 1.1* (52 g) was further purified by CC ( $\text{SiO}_2$ ; petroleum ether (PE)/AcOEt 2:1) to yield *Fr. 1.1.1* – *Fr. 1.1.4*. *Fr. 1.1.1* (29 g) was purified by CC ( $\text{SiO}_2$ ;  $\text{CHCl}_3/\text{Me}_2\text{CO}$  6:1) to provide *Fr. 1.1.1.1* – *Fr. 1.1.1.3*. *Fr. 1.1.1.1* (500 mg) was repeatedly subjected to RP-CC (1.  $C_{18}$ ,  $\text{MeOH}/\text{H}_2\text{O}$  50:50  $\rightarrow$  100:0; 2. *Sephadex LH-20*,  $\text{CHCl}_3/\text{MeOH}$  6:4) to afford **1** (13 mg) and **2** (6 mg). *Fr. 1.1.1.2* (1.2 g) was subjected to repeated vacuum liquid chromatography (VLC) ( $\text{SiO}_2$ ;  $\text{CHCl}_3/\text{i-PrOH}$  15:1) to yield **3** (107 mg). *Fr. 1.1.1.3* (300 mg) was also purified by VLC ( $\text{SiO}_2$ ; PE/i-PrOH 15:1) and then by RP-CC (*Sephadex LH-20*;  $\text{CHCl}_3/\text{MeOH}$  6:4) to yield **4** (26 mg) and **5** (22 mg).

**3,12,15-Trihydroxy-16,17-dimethoxytricyclo[12.3.1.1<sup>2,6</sup>]nonadeca-1(18),2(19),3,5,14,16-hexaene-9-one (1).** Colorless powder. UV (MeOH): 295, 258.  $[\alpha]_{\text{D}}^{23.5} = 0$  ( $c = 0.12$ , acetone).  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR: see Table. EI-MS: 372 ( $M^+$ ). HR-ESI-MS: 395.1467 ( $[M + \text{Na}]^+$ ,  $\text{C}_{21}\text{H}_{24}\text{NaO}_6^+$ ; calc. 395.1471).

**17-Methoxy-2-oxatricyclo[13.2.2.1<sup>3,7</sup>]jicosa-1(17),3(20),4,6,15,18-hexaene-4,10,11-triol (2).** Colorless powder. UV (MeOH): 279.  $[\alpha]_{\text{D}}^{23.6} = -71.4$  ( $c = 0.07$ , acetone).  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR: see Table. EI-MS: 344 ( $M^+$ ). HR-ESI-MS: 367.1517 ( $[M + \text{Na}]^+$ ,  $\text{C}_{20}\text{H}_{24}\text{NaO}_5^+$ ; calc. 367.1521).

### REFERENCES

- [1] K. R. Kuang, P. Q. Li, ‘Flora of China’, Science Publishing House, Beijing, 1979, Vol. 21, p. 6.
- [2] Yunnan Corporation of Materia Medica, ‘Yunnan Zhongyao Ziyuan Minglu’, Science Publishing House, Beijing, 1993, p. 68.
- [3] N. Sakurai, Y. Yaguchi, T. Inoue, *Phytochemistry* **1987**, 26, 217.
- [4] H. Matsuda, T. Morikawa, J. Tao, K. Ueda, M. Yoshikawa, *Chem. Pharm. Bull.* **2002**, 50, 208.
- [5] N. Masahiro, S. Nobuko, Y. Nobuyuki, N. Seiji, S. Shujiro, *Chem. Pharm. Bull.* **2000**, 48, 1427.
- [6] D. Sun, Z. Zhao, H. Wong, L. Y. Foo, *Phytochemistry* **1988**, 27, 579.

- [7] J. Tao, T. Morikawa, I. Toguchida, S. Ando, H. Matsuda, M. Yoshikawa, *Bioorg. Med. Chem.* **2002**, *10*, 4005.
- [8] K. E. Malterud, T. Anthonsen, *Phytochemistry* **1980**, *19*, 705.
- [9] B. S. Joshi, S. W. Pelletier, *J. Nat. Prod.* **1996**, *59*, 759.
- [10] M. Morihara, N. Sakurai, T. Inoue, K. Kawai, M. Nagai, *Chem. Pharm. Bull.* **1997**, *45*, 820.
- [11] M. Tene, H. K. Wabo, P. Kamnaing, A. Tsopmo, P. Tane, J. F. Ayafor, O. Sterner, *Phytochemistry* **2000**, *54*, 975.
- [12] Z. H. Zhou, C. R. Yang, *Yunnan Zhiwu Yanjiu* **2000**, *22*, 219.
- [13] K. E. Malterud, T. Anthonsen, J. Hjortas, *Tetrahedron Lett.* **1976**, *35*, 1674.
- [14] M. Kubo, T. Inoue, M. Nagai, *Chem. Pharm. Bull.* **1980**, *28*, 1300.
- [15] Y. Takeda, T. Fujita, T. Shingu, C. Ogimi, *Chem. Pharm. Bull.* **1987**, *35*, 2569.

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